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# Diagnostic value of IL-6 for patients with asthma: a meta-analysis



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# Abstract

REVIEW

**Background** IL-6 is a pleotropic cytokine that acts as a pro-inflammatory mediator and acute-phase response inducer, but has also been reported to possess anti-inflammatory properties. The objective of this study was to assess the validity of serum IL-6 test for diagnosis of asthma.

**Methods** A literature search was conducted using PubMed, Embase, and Cochrane library from January 2007 to March 2021 to identify relevant studies. Eleven studies were included in this analysis, involving 1977 patients with asthma and 1591 healthy non-asthmatic controls. The meta-analysis was performed using Review Manager 5.3 software and Stata 16.0. Random effect model or fixed effect model (FEM) was used to estimate the standardized mean differences (SMDs) with 95% confidence intervals (Cls).

**Results** The meta-analysis results revealed that the serum IL-6 levels were higher in asthmatic patients than healthy non-asthmatic controls (SMD 1.31, 95% CI 0.82–1.81, P < 0.00001). IL-6 levels are significantly elevated in pediatric patients with asthma (SMD 1.58, 95% CI 0.75–2.41, P = 0.0002) and mildly elevated in adult patients with asthma (SMD 1.08, 95% CI 0.27–1.90, P = 0.009). In addition, a subgroup analysis of asthma disease status showed that IL-6 levels were increased in stable (SMD 0.69, 95% CI 0.28–1.09, P=0.009) and exacerbation asthma (SMD 2.15, 95% CI 1.79–2.52, P < 0.00001) patients.

**Conclusion** The results of this meta-analysis suggest that serum IL-6 levels were significantly elevated in asthmatic patients as compared to normal population. IL-6 levels can be used as an auxiliary indicator to distinguish individuals with asthma from healthy non-asthmatic controls.

Keywords IL-6, Biomarker, Asthma, Meta-analysis

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# Introduction

Asthma is defined as reversible airflow obstruction in the setting of airway inflammation and is characterized by variable symptoms such as wheezing, breathlessness, chest tightness, and cough [1, 2]. Currently, asthma remains an important worldwide health problem in terms of both prevalence and severity in all regions and age group. There are an estimated 235–334 million asthma sufferers worldwide and it is responsible for approximately 250,000 deaths annually [3].

The extent of asthma is usually determined by a severity classification termed to be either intermittent or



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persistent. Persistent asthma then is classified as mild, moderate, or severe. In addition, asthma is a heterogeneous disease encompassing different phenotypes. asthma patients can be classified into allergic, nonallergic, occupational, aspirin-exacerbated respiratory disease, potentially fatal, exercise-induced, and cough variant asthma [4]. Traditionally asthma diagnosis is predominantly based on the combined of clinical history, presence of typical symptoms, and objective tests of lung function or airway hyperresponsiveness [5]. However, there are some patients who failed to get the lung function and airway hyperresponsiveness objective measurement, which may lead to inappropriate underdiagnosis. Therefore, it is important to find new potential asthma biomarkers, which could contribute to a better understanding of the pathophysiological mechanisms of asthma and to the search for new therapeutic targets.

Specific biomarkers may guide diagnosis, treatment, and predict treatment responses [6]. IL-6 is a typical cytokine with roles in the immune response, inflammation, hematopoiesis and in the endocrine and nervous systems [7]. In early 1980, a study found that there was an active factor that could induce immunoglobulin (Ig) production in Epstein-Barr virus (EBV) -transformed B lymphoblastoid cell lines [8, 9]. Furthermore, the plasmacytoma/ hybridoma/myeloma growth factor and the hepatocyte stimulating factor were also found to be identical to this factor [10, 11]. The factor was later named as "interleukin 6" [12]. IL-6 is produced by cells of the innate immune system, B cells, and a small number of CD4 effector Th cells. In addition, IL-6 is also secreted by non-leukocytes such as endothelial cells, fibroblasts, astrocytes, epithelial cells and some malignant cells [13]. These cells can be stimulated to produce IL-6 by actions that promote cell stress or damage (such as UV, irradiation, ROS, microbial products, viruses, or other pro-inflammatory cytokines) [14]. IL-6, which is rapidly and transiently produced in response to infections and tissue injurys, promotes host defense by stimulating acute phase responses, hematopoiesis, and immune responses. Therefore, IL-6 has long been considered a general marker of inflammation. However, IL-6 is now found to be a marker of certain inflammatory diseases. The last decade or so has seen an increasing number of studies shedding light on IL-6 as key cell signaling mode in asthma-related pathways and may play a role as a biomarker of asthma [15]. However, most of these studies have small sample sizes, and a single study may lack sufficient statistical relevance to detect the potentially subtle effects of the IL-6 levels on asthma. Thus, we performed a meta-analysis to accurately investigate the association between IL-6 and asthma.

# Methods

# Search strategy

We searched several commonly used databases (from January 1, 2007 until March 8, 2021) from PubMed, Embase, and the Cochrane Library. A combination of searching terms were used to search those three electronic databases, relating to the following two concepts: (1) IL-6 ('Interleukin-6' OR ' B cell stimulatory factor 2' OR 'B cell differentiation factor 2' OR 'BSF-2'OR'IL6'OR 'hybridoma growth factor' OR 'plasmacytoma growth factor 'OR 'hepatocyte stimulating factor' OR 'MGI-2' OR' myeloid differentiation inducing protein 'OR 'interferon beta 2') and (2) asthma ('Asthma' OR' bronchial asthma' OR 'Asthmas' OR 'asthma bronchial'). The searches were restricted to studies of only human subjects, and only articles in the English and Chinese language were applied. The references of included studies or reviews were also checked for additional reports.

### Study selection

The inclusion criteria were defined as follows: (1) patients with asthma were studied, (2) express IL-6 levels as mean $\pm$ standard deviation, median and range, or median and interquartile range, (3) include patients with asthma and controls, who had no history of asthma or other respiratory disease, (4) if there was duplication of data, only the most complete and recent study was included, and (5) the study design is a case-control study.

### Data extraction and quality assessment

The general characteristics of the study were extracted using a standardized data extraction form: publication information (first author's name, Publication year), study population, sample size, IL-6 measurement, and IL-6 levels and units(ng/L) in patients with asthma and controls. In terms of quality evaluation, we applied the Newcastle–Ottawa Quality Assessment Scale, with a total score of 0–3, 4–6, and 7–9 considered low, moderate, and high quality, respectively.

## Statistical analysis

The meta-analysis was performed using Review Manager 5.3 software and Stata 16.0. The means and standard deviation were pooled and calculated between patients with asthma and controls. Wood et al. reported median and interquartile range, we analyzed the mean and standard deviation mathematically [16, 17]. In addition, we analyzed the age of patients with asthma, including adults and children with asthma, as well as the status of asthmatic patients (asthma exacerbation and stable asthma). Exacerbation of asthma was defined as the sudden onset of symptoms, such as wheezing, shortness of breath, cough and chest tightness, or a worsening of the original symptoms, often with dyspnea, characterized by a decrease in forced expiratory volume in one second (FEV1). Stable asthma was defined as stable symptoms and lung function maintained for at least 4 weeks. Heterogeneity was assessed using a chi-squared Q test and I-squared statistics. If PQ<0.1 or I  $^2$ > 50%, the heterogeneity was considered significant, and a random effects model (REM) was used otherwise a fixed effect model (FEM) was used. If the mean level differences were significant across studies, or different units were used, standardized mean difference (SMD) was used to estimate the effect size. In the included studies, the serum IL-6 were measured in ELISA with different source reagent, and the differences in the mean levels of IL-6 were considered significant, therefore, SMD in serum IL-6 was used to estimate the effect size. Publication bias was assessed by examining the funnel plot. A sensitivity analysis was performed to explore the stability of the meta-analysis.

# Results

# The general data

The main characteristics of the included studies are summarized in Table 1 and the steps for screening and the study selection procedure are presented in Fig. 1 .We initially identified 1385 records in the electronic databases and after removing duplicates, 1242 records were left for title and abstract screening. Finally, we obtained a total of eleven relevant studies [18–28] on IL-6, which consist of 1977 patients with asthma and 1591 healthy non-asthmatic controls.

Note that enzyme-linked immunosorbent assay (ELISA) was used in all included studies. According to the Newcastle–Ottawa Scale, eleven studies had high quality. Low-quality studies were not included in this meta-analysis.

# Correlation of IL-6 in patients with asthma

Substantial heterogeneity was observed ( $I^2=96\%$ ) among studies. Thus, our analysis was performed using a

random-effects model. Figure 2 provided a comparison of serum IL-6 levels between asthma patients and controls in the included studies. The pooled SMDs revealed that serum IL-6 levels were significantly higher in the asthma group as compared to the healthy non-asthmatic controls group (SMD=1.31, 95% CI: 0.82-1.81, P<0.00001).

# Sensitivity analysis and publication bias

The funnel plots were substantially asymmetric (**Supplementary Fig. 1**), indicating the presence of a possible publication bias. **Supplementary Fig. 2** showed estimates in the random effects model, with the middle line representing the calculated median (2.01) for all samples, the left and right lines representing the lower (1.40) and higher (3.58) of 95% confidence intervals, respectively, and each circle representing the new mean value obtained after removing a given study from the pool. This indicated that all data were within the 95% confidence intervals for the total data set and that no single study had a large effect on the mean value.

## Subgroup analysis

Based on the age of patients (Children/Adult), we conducted subgroup analyses to evaluate the association between the age of patients and IL-6 levels. As in a subgroup of adult asthma, six studies [18–23] were included and provided a total number of 458 participants. The estimated SMD (1.08, 95% CI 0.27–1.90, P=0.009) indicated that serum IL-6 levels were higher in asthmatics as compared to control group (Fig. 3).Five studies [24–28]were included in the subgroup of children with asthma, contributing 1,519 participants for this meta-analysis. The estimated SMD of 1.58 (95% CI 0.75–2.41, P<0.0002) showed that serum IL-6 levels were higher than those in the control group (Fig. 4).

Considering the asthma status ( asthma exacerbation/ stable asthma), the results of subgroup analysis of asthma exacerbation showed that serum IL-6 levels

Table 1 The general data of included documents

Study Year Samples Country Size(A/C) Sex(M/F) Method Asthma Control Ding [18] 2015 Serum China 120/120 62/58 60/60 FLISA Radulovic [19] 2015 12/10 9/3 10/ NA ELISA Serum America Canöz [20] 2008 Serum Turkey 54/42 NA /54 NR ELISA Naik [21] 2017 India 14/16 ELISA Serum 50/3027/23 Domvri [22] 2019 Serum Greece 90/30 31/59 10/20 ELISA Wood [23] 2012 Serum Australia 132/83 55/77 31/52 ELISA Ma [24] 2019 China 192/130 97/95 67/63 ELISA Serum Zhang [25] 2018 China 70/25 ELISA Serum 25/45 8/15 Cui [26] 2017 719/439 ELISA Serum China 1158/1075 686/389 Fabian [27] 2011 Serum Hungary 35/21 28/7 16/5 ELISA Cui [28] 2014 Serum China 64/25NR NR ELISA

Abbreviations: ELISA, enzyme-linked immunosorbent assay; A, asthma; C, control; M, male; F, female; NR, not report; NA, not applicable

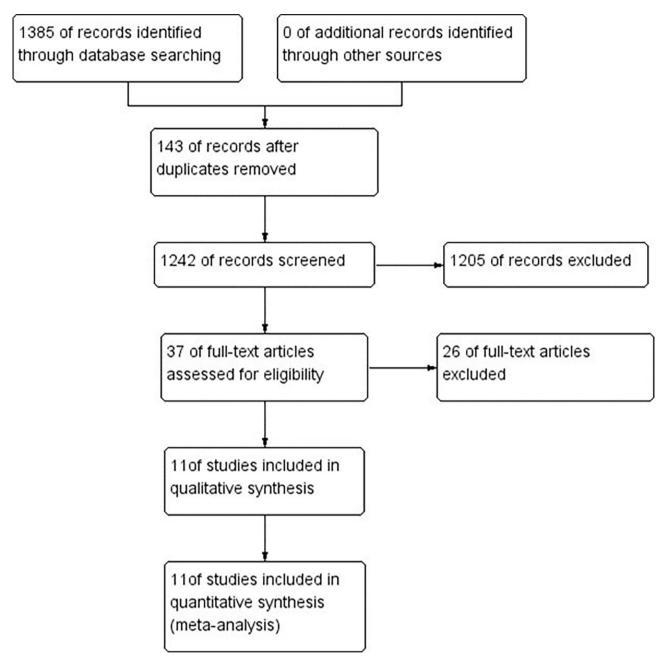


Fig. 1 Flow chart of searching the relevant studies used in this meta-analysis

were significantly higher in patients with asthma exacerbation than in healthy non-asthmatic controls, with a pooled SMD of 2.15 (95% CI 1.79–2.52, P<0.00001)[18, 28] (Fig. 5). The similar results were also found in stable asthma (SMD 0.69, 95% CI 0.28–1.09, P=0.0009)[18, 23, 28] (Fig. 6).

## **Meta-regression analysis**

Due to the high heterogeneity in Fig. 2, we conducted multivariate meta-regression analysis to study the confounding factors that may lead to this phenomenon. The results showed that year, country and age were not the source of the heterogeneity, which may be caused by multiple factors.

# Discussion

The present study was conducted to investigate the potential role of IL-6 in asthma patients. The main findings of this meta-analysis are (1) patients with asthma had higher serum IL-6 concentrations than those in controls; (2) Serum IL-6 levels are elevated in both adult and pediatric asthmatics compared to controls, especially in pediatric asthmatics; (3) Serum IL-6 levels were higher

	A	sthma		0	Control			Std. Mean Difference	Std. Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% Cl	IV, Random, 95% Cl
Canöz 2008[20]	24.83	4.17	54	7.46	4.06	42	8.2%	4.18 [3.45, 4.91]	
Cui2014[28]	17.72	5.55	64	10.2	2.5	25	9.0%	1.52 [1.01, 2.04]	+
Cui2017[26]	19.31	11.6	1158	10.23	4.62	1075	10.0%	1.01 [0.93, 1.10]	•
Ding2015[18]	38.41	18.44	120	26.3	11.5	120	9.7%	0.79 [0.52, 1.05]	*
Domvri2019[22]	86	266	90	7	4	30	9.3%	0.34 [-0.08, 0.76]	+
Fabian2011[27]	0.158	0.053	35	0.099	0.066	21	8.8%	1.00 [0.43, 1.57]	
Ma2019[24]	7.5	1.48	192	3.95	0.72	130	9.6%	2.88 [2.56, 3.19]	+
Naik2017[21]	54.63	23.05	50	25.53	3.93	30	9.0%	1.56 [1.05, 2.08]	
Radulovic2015[19]	7.6	2.1	12	54.7	68.9	10	7.5%	-0.98 [-1.88, -0.08]	
Wood2012[23]	1.65	0.97	132	1.14	0.53	83	9.7%	0.61 [0.33, 0.89]	+
Zhang2018[25]	4.45	2.19	70	1.58	1.03	25	9.1%	1.45 [0.95, 1.96]	+
Total (95% CI)			1977			1591	100.0%	1.31 [0.82, 1.81]	•
Heterogeneity: Tau <sup>2</sup> :	= 0.64; C	hi <sup>2</sup> = 25	2.71, d	f = 10 (F	< 0.00	001); F	= 96%		
Test for overall effect: $Z = 5.20$ (P < 0.00001)									-10 -5 0 5 10 Favours [experimental] Favours [control]

Fig. 2 Comparison of serum IL-6 levels between asthma patients and healthy non-asthmatic controls in the included studies

	Adu	It asthn	na	C	ontrol		5	Std. Mean Difference	Std. Mean Difference				
Study or Subgroup	Mean SD		Total	Mean	SD	Total	Weight	IV, Random, 95% Cl	IV, Random, 95% Cl				
Canöz 2008[20]	24.83	4.17	54	7.46	4.06	42	15.8%	4.18 [3.45, 4.91]	+				
Ding2015[18]	38.41	18.44	120	26.3	11.5	120	17.7%	0.79 [0.52, 1.05]	•				
Domvri2019[22]	86	266	90	7	4	30	17.2%	0.34 [-0.08, 0.76]	+				
Naik2017[21]	54.63	23.05	50	25.53	3.93	30	16.8%	1.56 [1.05, 2.08]					
Radulovic2015[19]	7.6	2.1	12	54.7	68.9	10	14.8%	-0.98 [-1.88, -0.08]					
Wood2012[23]	1.65	0.97	132	1.14	0.53	83	17.7%	0.61 [0.33, 0.89]	•				
Total (95% CI)			458			315	100.0%	1.08 [0.27, 1.90]	•				
Heterogeneity: Tau <sup>2</sup> =	= 0.96; C	hi <sup>2</sup> = 11	2.86, d	f= 5 (P	< 0.00	001); F	= 96%	-					
Test for overall effect	: Z = 2.60	) (P = 0.	009)						-10 -5 0 5 10 Favours [experimental] Favours [control]				

Fig. 3 Forest plot of comparison serum IL-6 levels of adult asthma versus the healthy non-asthmatic controls

	Childh	ood astl	nma	Control				Std. Mean Difference	Std. Mean Difference
Study or Subgroup	Mean SD		Total	Mean	SD	Total	Weight	IV, Random, 95% Cl	IV, Random, 95% Cl
Cui2014[28]	17.72	5.55	64	10.2	2.5	25	19.6%	1.52 [1.01, 2.04]	*
Cui2017[26]	19.31	11.6	1158	10.23	4.62	1075	21.1%	1.01 [0.93, 1.10]	
Fabian2011[27]	0.158	0.053	35	0.099	0.066	21	19.2%	1.00 [0.43, 1.57]	-
Ma2019[24]	7.5	1.48	192	3.95	0.72	130	20.5%	2.88 [2.56, 3.19]	•
Zhang2018[25]	4.45	2.19	70	1.58	1.03	25	19.6%	1.45 [0.95, 1.96]	*
Total (95% CI)			1519			1276	100.0%	1.58 [0.75, 2.41]	•
Heterogeneity: Tau <sup>2</sup> =	= 0.85; Ch	i <sup>2</sup> = 127.	63, df =	4 (P < (	0.00001	); I <sup>2</sup> = 9	7%		
Test for overall effect	Z = 3.72	(P = 0.00	002)						-10 -5 0 5 10 Favours [experimental] Favours [control]

Fig. 4 Forest plot of comparison serum IL-6 levels of childhood asthma versus the healthy non-asthmatic controls

	Exacerba	tion of ast	thma	Control			St	d. Mean Difference	Std. Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Fixed, 95% CI	IV, Fixed, 95% Cl
Cui2014[28]	18.2	4.5	36	10.2	2.5	25	33.0%	2.07 [1.43, 2.71]	
Ding2015[18]	56.4	19.3	36	26.3	11.5	120	67.0%	2.19 [1.74, 2.64]	•
Total (95% CI)			72			145	100.0%	2.15 [1.79, 2.52]	+
Heterogeneity: Chi2 =	0.09, df = 1	(P = 0.76);	$ ^{2} = 0\%$						-20 -10 0 10 20
Test for overall effect:	Z=11.54 (P	< 0.00001	1)						Favours [experimental] Favours [control]

Fig. 5 Forest plot of comparison serum levels of IL-6 in the asthma exacerbation group versus the healthy non-asthmatic controls

in patients with exacerbation and stable asthma than in healthy non-asthmatic controls.

IL-6 levels usually rise when cells are stressed or injured. The researchers found that serum IL-6 levels were elevated, which was related to inflammatory

diseases [29]. It is acknowledged that airway inflammation is a prominent feature of asthma [30]. Interestingly, lung epithelial cells can secrete cytokines such as IL-6 and regulate immunity [31]. It has also been reported that IL-6 is overexpressed in bronchial epithelial cells in adults

	Stabl	e asth	ma	Control				Std. Mean Difference	Std. Mean Difference				
Study or Subgroup	Mean SD Total			Mean SD		Total	Weight	IV, Random, 95% Cl		IV, Ra	andom, 95	5% CI	
Cui2014[28]	17.1	6.7	28	10.2	2.5	25	23.2%	1.32 [0.72, 1.91]			-		
Ding2015[18]	30.7	11.4	84	26.3	11.5	120	38.4%	0.38 [0.10, 0.66]					
Wood2012[23]	1.65	0.97	132	1.14	0.53	83	38.4%	0.61 [0.33, 0.89]			-		
Total (95% CI)			244			228	100.0%	0.69 [0.28, 1.09]			٠		
Heterogeneity: Tau <sup>2</sup> =					0.02);	<sup>2</sup> = 749	6		-10	-5	-	5	10
Test for overall effect	: Z = 3.32	P = 0	.0009)						Favours [	-	ntal) Favo	ours [cont	

Fig. 6 Forest plot of comparison serum levels of IL-6 in the stable asthma group versus the healthy non-asthmatic controls

and children with asthma [32]. In addition, Asthma is treated with inhaled corticosteroids which shut down the expression of asthma-related cytokines such as IL-6[33]. Taken together, these results suggest that serum IL-6 levels may be elevated in asthmatic patients. There is growing evidence that IL-6 may also play an important role in the initial development and subsequent progression of asthma [34, 35].

In present meta-analysis, we observed significant heterogeneity in these studies. This heterogeneity remained after the subgroup analysis, suggesting that the study population, measure reagents, age and other covariates may be responsible for it. Therefore, we performed a meta-regression analysis to explore possible sources of heterogeneity, and the results showed that none of these factors was a source of heterogeneity. The ELISA kits in the study were purchased from different manufacturers. Thus, among the included studies, the sensitivity of the measurement reagents was different, which can be a possible factor for heterogeneity. Furthermore, Sensitivity analyses indicated that the pooled SMD were not excessively affected by a single study.

This paper has several limitations. First of all, only articles in the English and Chinese language studies are picked in this meta-analysis, and articles in other languages that meet the inclusion criteria may be missed, which may lead to publication bias. Secondly, the sample size of the eleven articles included in this study is small, which may be one of the reasons to explain the existence of publication bias.

# Conclusion

In summary, although limited studies have reported serum IL-6 level in asthma, our findings associate higher levels of IL-6 in patients with asthma, especially in children with asthma. Thus, IL-6 levels can be used as an auxiliary indicator to distinguish individuals with asthma from healthy non-asthmatic controls.

## **Supplementary Information**

The online version contains supplementary material available at https://doi. org/10.1186/s13223-023-00794-3.

 $\label{eq:supplementary Figure 1. Funnel plot with pseudo 95\% \ confidence intervals$ 

Supplementary Figure 2. Meta-analysis random-effects estimates for all the included studies

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# Authors' contribution

RP and SK were responsible for methodology, data analysis. QL and XZ contributed to data analysis. RP, TW and YC contributed to the writing idea, data analysis and original writing. YC supervised and contributed to data analysis and writing. All authors have read and approved the final manuscript.

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#### Data availability

The datasets used and analyzed during the current study can available from the corresponding author on reasonable request.

# Declarations

#### Ethics approval and consent to participate

The study does not require ethical approval because the mata-analysis is based on published research and the original data are anonymous.

#### **Consent for publication**

Authors are the sole responsible for the publication of this study.

#### **Competing interest**

Authors have no competing interests to declare.

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# References

- Erzurum SC, Gaston BM. Biomarkers in Asthma. Clin Chest Med. 2012;33(3):459–71.
- Pearce N, Ait-Khaled N, Beasley R, et al. Worldwide trends in the prevalence of asthma symptoms: phase III of the International Study of Asthma and Allergies in Childhood (ISAAC). Thorax. 2007;62(9):758–66.
- Kim H, Ellis AK, Fischer D, et al. Asthma biomarkers in the age of biologics. Allergy Asthma Clin Immunol. 2017;13:48. https://doi.org/10.1186/ s13223-017-0219-4.
- Padem N, Saltoun C. Classification of asthma. Allergy Asthma Proc. 2019;40(6):385–8.
- Saglani S, Menzie-Gow AN. Approaches to Asthma diagnosis in children and adults. Front Pediatr. 2019;7:148.
- Licari A, Castagnoli R, Brambilla I, et al. Asthma endotyping and biomarkers in Childhood Asthma. Pediatr Allergy Immunol Pulmonol. 2018;31(2):44–55.

- Kaur S, Bansal Y, Kumar R, et al. A panoramic review of IL-6: structure, pathophysiological roles and inhibitors. Bioorg Med Chem. 2020;28:115327.
- Muraguchi A, Kishimoto T, Miki Y, et al. T cell-replacing factor- (TRF) induced IgG secretion in a human B blastoid cell line and demonstration of acceptors for TRF. J Immunol. 1981;127(2):412.
- Teranishi T, Hirano T, Arima N, et al. Human helper T cell factor(s) (ThF). II. Induction of IgG production in B lymphoblastoid cell lines and identification of T cell-replacing factor- (TRF) like factor(s). J Immunol. 1982;128(4):1903.
- Van Damme J, Opdenakker G, Simpson RJ, et al. Identification of the human 26-kD protein, interferon beta 2 (IFN-beta 2), as a B cell hybridoma/plasmacytoma growth factor induced by interleukin 1 and tumor necrosis factor. J Exp Med. 1987;165(3):914–9.
- Van Snick J, Cayphas S, Szikora JP, et al. cDNA cloning of murine interleukin-HP1: homology with human interleukin 6. Eur J Immunol. 1988;18(2):193–7.
- Sehgal PB, Helfgott DC, Santhanam U, et al. Regulation of the acute phase and immune responses in viral disease. Enhanced expression of the beta 2-interferon/hepatocyte-stimulating factor/interleukin 6 gene in virusinfected human fibroblasts. J Exp Med. 1988;167(6):1951–6.
- 13. Hirano T. Interleukin 6 and its receptor: ten years later. Int Rev Immunol. 1998;16(3–4):249–84.
- 14. Rincon M, Irvin CG. Role of IL-6 in asthma and other inflammatory pulmonary diseases. Int J Biol Sci. 2012;8(9):1281–90.
- Poynter ME, Irvin CG. Interleukin-6 as a biomarker for asthma: hype or is there something else? Eur Respir J. 2016;48(4):979–81.
- Luo D, Wan X, Liu J, et al. Optimally estimating the sample mean from the sample size, median, mid-range, and/or mid-quartile range. Stat Methods Med Res. 2018;27(6):1785–805. https://doi.org/10.1177/0962280216669183.
- Wan X, Wang W, Liu J, et al. Estimating the sample mean and standard deviation from the sample size, median, range and/or interquartile range. Bmc Med Res Methodol. 2014;14:135.
- Ding Y, Yang H, He H, et al. Plasma adiponectin concentrations and adiponectin gene polymorphisms are Associated with Bronchial Asthma in the Chinese Li Population. Iran J Allergy Asthma Immunol. 2015;14(3):292–7.
- 19. Radulovic M, Bauman WA, Wecht JM, et al. Biomarkers of inflammation in persons with chronic tetraplegia. J Breath Res. 2015;9(3):036001.
- Canöz M, Erdenen F, Uzun H, et al. The relationship of inflammatory cytokines with asthma and obesity. Clin Invest Med. 2008;31(6):E373–9.
- 21. Naik SP. Evaluation of inflammatory markers interleukin-6 (IL-6) and matrix metalloproteinase-9 (MMP-9) in asthma. J Asthma. 2017;54(6):584–93.
- Domvri K, Porpodis K, Tzimagiorgis G, et al. Th2/Th17 cytokine profile in phenotyped greek asthmatics and relationship to biomarkers of inflammation. Respir Med. 2019;151:102–10.

- Wood LG, Baines KJ, Fu J, et al. The neutrophilic inflammatory phenotype is associated with systemic inflammation in asthma. Chest. 2012;142(1):86–93.
- 24. Ma SL, Zhang L. Elevated serum OX40L is a biomarker for identifying corticosteroid resistance in pediatric asthmatic patients. Bmc Pulm Med. 2019;19(1):66.
- 25. Zhang Y, Li T, Yuan H, Pan W, Dai Q. Correlations of inflammatory factors with intestinal Flora and gastrointestinal incommensurate symptoms in children with asthma. Med Sci Monit. 2018;24:7975–9.
- Cui AH, Zhao J, Liu SX, et al. Associations of IL-4, IL-6, and IL-12 levels in peripheral blood with lung function, cellular immune function, and quality of life in children with moderate-to-severe asthma. Med (Baltim). 2017;96(12):e6265.
- Fabian E, Pölöskey P, Kósa L, et al. Activities of antioxidant enzymes in relation to oxidative and nitrosative challenges in childhood asthma. J Asthma. 2011;48(4):351–7. https://doi.org/10.3109/02770903.2011.560319.
- Cui YJ, Chen GH, Wang JL, et al. [Alterations of CD4 + CXCR5 + tfh cells and its transcription regulatory factors in children with asthma]. Zhongguo Dang Dai Er Ke Za Zhi. 2014;16(12):1215–9.
- Kishimoto T. IL-6: from its discovery to clinical applications. Int Immunol. 2010;22(5):347–52.
- 30. Stephen T, Holgate et al. Asthma Nat Rev Dis Primers 2015.
- Crestani B, Cornillet P, Dehoux M, et al. Alveolar type II epithelial cells produce interleukin-6 in vitro and in vivo. Regulation by alveolar macrophage secretory products. J Clin Invest. 1994;94(2):731–40.
- Marini M, Vittori E, Hollemborg J, et al. Expression of the potent inflammatory cytokines, granulocyte-macrophage-colony-stimulating factor and interleukin-6 and interleukin-8, in bronchial epithelial cells of patients with asthma. J Allergy Clin Immunol. 1992;89(5):1001–9.
- Levine SJ, Larivée P, Logun C, et al. Corticosteroids differentially regulate secretion of IL-6, IL-8, and G-CSF by a human bronchial epithelial cell line. Am J Physiol. 1993;265(4 Pt 1):L360–8.
- Neveu WA, Allard JB, Dienz O, et al. IL-6 is required for airway mucus production induced by inhaled fungal allergens. J Immunol. 2009;183(3):1732–8.
- Neveu WA, Bernardo E, Allard JL, et al. Fungal allergen β-glucans trigger p38 mitogen-activated protein kinase-mediated IL-6 translation in lung epithelial cells. Am J Respir Cell Mol Biol. 2011;45(6):1133–41.

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